

# PATsmart™ ZipChip®

## Essential Procedures

SW version 1.5.1.7 and above

### Initialize ZipChip

1. Before starting, ensure that your mass spectrometer is calibrated, and that ion transfer tube/capillary were recently cleaned/changed.
2. Remove current mass spec source and disconnect any contact closure cables. Remove sweep cone if used. Attach the ZipChip Interface and its contact closure cable(s) to the mass spec. (*for more details see 908devices.com/support*)
3. Connect USB cable, contact closure/autosampler cable, power cable, waste tubing, and BGE & sample transfer lines to the ZipChip interface.
  - a. Tighten transfer lines until resistance is felt, then add another quarter turn.
4. Open the ZipChip software.
5. From the Setup Tab:
  - a. Perform an **Autosampler Prime** using appropriate BGE.
  - b. Perform a **Chip Prime** using appropriate chip type.

### Resuming After Long Batches & Idling

1. Replace chip with Manifold Dryer Chip.
2. Perform *Dry Manifold* from the ZipChip software.
3. Replace the chip. Open the ZipChip software and perform:
  - a. *BGE Refresh*.
  - b. *Sample Well Rinse*.
  - c. *Infusion* (0 V/cm) for 4 minutes.

### Shutdown Procedures

1. Perform a **BGE Refresh** from the Setup Tab.
2. From the Singles Tab, Perform a **Sample Well Rinse**, then **Empty Sample Well**.
3. Remove the chip and empty all wells using a pipette.
4. Place chip into Dry Dock loosely, and then:
  - a. Turn on the gas but do not press down on the chip. Dry for 3 minutes before turning off gas.
  - b. Inspect chip wells for any microdroplets.
  - c. If droplet-free, reinsert the chip into Dry Dock, and this time, push down firmly and lock the hatch. Turn on gas. If droplets are visible, repeat steps 4a-b.
  - d. Dry for a minimum of 30 minutes.
5. Perform a **Dry Manifold** from the Setup Tab
6. Replace the Manifold Dryer Chip with a blank chip.
7. Close the ZipChip Software.
8. If changing to another MS source, disconnect the ZipChip interface. If possible, leave Sample & BGE transfer line connections intact. Place ZipChip on top of Autosampler.

### Recommended Consumables & Settings

#### Charge Variant Analysis (CVA)

##### CVA Kit • HRN Chip

Sample Conc: 0.1–1 mg/mL diluted in CVA Diluent  
 Injection Volume: 1 nL • Analysis Time: 10–15 mins  
 BGE Refresh Rate: Every 1 line

#### Intact Denatured Analysis

##### Peptides Kit • HS or HR Chip

Sample Conc: 0.1–1 mg/mL diluted in Peptides BGE  
 Injection Volume: 1 nL • Analysis Time: 1–5 mins BGE  
 Refresh Rate: Every 1 line

#### Subunit Analysis

##### Peptides Kit • HR Chip

Sample Conc: 0.1–1 mg/mL diluted in Peptides BGE  
 Injection Volume: 1 nL • Analysis Time: 4–10 mins  
 BGE Refresh Rate: Every 1 line

#### Peptide Mapping

##### Peptides Kit • HR Chip

Sample Conc: 0.5 mg/mL diluted in Peptides Diluent  
 Injection Volume: 5.5 nL • Analysis Time: 7–15 mins  
 BGE Refresh Rate: Every 2–3 lines

#### Small Molecules, AAA, and Metabolomics

##### Peptides Kit • HR Chip

Sample Conc: 10 µM diluted in Peptides Diluent  
 Injection Volume: 5.5 nL • Analysis Time: 5–10 mins  
 BGE Refresh Rate: Every 2–3 lines

#### Oligonucleotide Analysis

##### Oligos Kit/Oligos Starter Kit • HSB or HRB Chip

Sample Conc: 1–10 µM diluted into Oligos Diluent  
 Injection Volume: 1 nL • Analysis Time: 2–10 mins  
 BGE Refresh Rate: Every 1 line

### Changing BGE

If changing from low-pH BGE (Peptides) to high-pH BGE (CVA or Oligos) follow the Neutral Quenching Protocol as outlined in each corresponding protocol.

### Consumables Handling & Storage

- Always wear gloves when handling chips. Never touch the exposed glass corner. Store all chips in an enclosed container. Minimize lab-air exposure of BGE.
- BGE, diluent, and chips can be stored at room temperature.
- Reagents have a 90-day working life after first use.